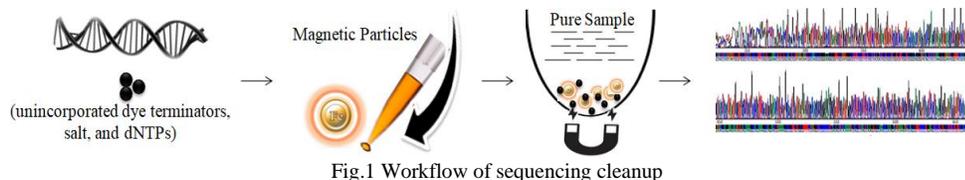


## One-Step Sequencing Cleanup Kit

Sanger sequencing, also known as "chain terminator sequencing," is a method of DNA sequencing that relies on DNA polymerase's selective incorporation of fluorescent-labeled (bigdye terminator) dideoxy nucleotide chain terminators during in vitro DNA replication for use in a single sequencing reaction. The chain termination PCR produces DNA fragments of varying lengths, each of which ends with a fluorescently labeled dideoxynucleotide. It functions similarly to standard PCR, with one key exception. In contrast to regular PCR, a low ratio of modified nucleotides is introduced along with normal dNTPs. These modified dNTPs are known as dideoxynucleotides (ddNTPs) and have a fluorescent label known as Dye. It quickly became the most used sequencing method for a variety of applications, including de novo sequencing, mutation discovery and confirmation, and resequencing.

After the cycle sequencing reaction, it is necessary to remove contaminants from DNA extension products (e.g., unincorporated fluorescent dyes, residual salts, dNTPs, primers, and enzymes). These impurities, such as dye peaks or "dye blobs," can frequently interfere with the quality and signal intensity of sequencing data, obscuring sections of the sequencing chromatogram and interfering with the base-calling accuracy of sequencing analysis tools. To clean and purify the sequencing reaction extension items on the market, a number of dye-terminator removal (sequencing clean up or sequencing purification) products are available. However, those sequencing purification protocols are either time-consuming (for example, using a spin column, ethanol precipitation, and SPRI paramagnetic beads) or result in sample loss due to protein precipitation, several transfer stages, and a lengthy process.

**BcMag™ One-Step Sequencing Cleanup Kit** is specifically designed for fast and efficient purification of the post-Sanger Sequencing reaction. The entire protocol takes only one tube and is complete in less than 5minutes (Fig 1). The magnetic beads are added directly to the finished sequencing reactions and vortexed to capture the impurities (e.g., unincorporated dyes, dNTPs, residual salts, and other interfering components). After vortexing, the beads are magnetically captured, while the clean supernatant can be directly loaded onto a capillary sequencer.



**Features and Advantages:**

- One tube, 3 min protocol, No sample loss
- Reliable results: excellent Long and short fragment recovery, Q20 read length > 800 bases
- Cost-Effective: Tremendously reduced labor costs and other consumed material such as columns, filters, laborious repeat pipetting, and ethanol.
- High-throughput: Compatible with many different automated liquid handling systems.
- Compatible with BigDye XTerminator run modules, e.g., unnecessary to remove the magnetic beads from the tube, the supernatant can be directly loaded onto the capillary sequencer.
- Efficient removal of any dye terminator

**Handling and Storage**

- Store at 4°C upon arrival for up to 6 months.

| Components                             | Storage | 100 preps, Cat # AI-101 | 250 preps, Cat # AI-102 |
|--|---------|-------------------------|-------------------------|
| BcMag™ One-Step Sequencing Cleanup Kit | 4°C     | 1ml                     | 5 ml                    |

**PROTOCOL**

**IMPORTANT!**



The following protocol is optimized for the efficient purification of sequencing reactions containing 2  $\mu$ l (1:4 dilution of the terminator) or less of BigDye Terminator v3.1 and v1.1 or other dye terminators in a total reaction volume of 10  $\mu$ l. If an alternative reaction scale or dye terminator is used, the procedure may need to be optimized.

There are two methods of purifying the DNA sequencing reaction products.

1. Using 96-well PCR plates/tubes
2. Using 96-Well Microplates.

### Materials Required by the User

| Item   | Source                                  |
|--|---|
| 18.2 M $\Omega$ .cm, DNase/RNase-Free Ultrapure Water  |   |
| BcMag™ 96-well Plate Magnetic Rack (side-pull) compatible with 96-well PCR plate and 96-well microplate or other compatible racks.   | Bioclone, Cat#: MS-06                   |
| Adjustable Single and Multichannel pipettes  |   |
| Centrifuge with swinging bucket  |   |
| Addition items are required if using 96-well PCR plates/tubes  |   |
| Vortex Mixer<br>** The user can also use other compatible vortex mixers. However, the time and Speed should be optimized, and the mixer should be Orbit $\geq$ 1.5 mm-4 mm, Speed $\geq$ 2000 rpm  |   |
| Eppendorf™ MixMate™  | Eppendorf, Cat#:5353000529              |
| Tube Holder PCR 96   | Eppendorf, Cat#: 022674005              |
| Smart Mixer, Multi Shaker  | BenchTop Lab Systems, Cat#:5353000529   |
| PCR plates/tubes<br>** <b>IMPORTANT!</b> If using other tubes or PCR plates, ensure that the well diameter at the bottom of the conical section of PCR Tubes or PCR plates has to be $\geq$ 2.5mm. |   |
| 0.2 ml 96-well PCR Plates  | Applied Biosystems, Cat #: 4306737      |
| Olympus 0.2 ml 96-Well PCR Plate, Non-Skirted  | Genesee Scientific, Cat #: 24-300       |
| 0.2 mL Olympus 8-Strip PCR Tubes   | Genesee Scientific, Cat #: 24-706)      |
| 0.2mL Axygen™ 8-Strip PCR Tubes  | Fisher, Cat #: 14-222-252)              |
| Addition items are required if using 96-well microplates   |   |
| Vortex Mixer<br>** The user can also use other compatible vortex mixers. However, the time and Speed should be optimized, and the mixer should be: Orbit $\geq$ 3 mm-4 mm, Speed $\geq$ 800 rpm    |   |
| Fisher Scientific™ Microplate Advanced Vortex Mixers   | Fisher, Cat#:02-216-101                 |
| OHAUS Microplate Vortex Mixers   | OHAUS, Cat#:30392160                    |
| <b>IMPORTANT!</b> If using 96-well microplates, choose clear Flat-bottom Non-Binding Assay Microplates.  |   |
| Thermo Scientific™ 96-Well Microtiter™ Microplates   | Fisher, Cat #: 14-245-142, or 14-245-71 |
| Greiner Bio-One 96-Well Non-Binding Microplates  | Fisher, Cat #: 07-000-090               |
| Eppendorf™ MTP 96 Microplates  | Eppendorf Cat #: 951040048              |

### Procedure

- A. Prepare the Premix Beads solution



**IMPORTANT!**

- The magnetic beads and water can be added as a premix or sequentially.
  - Do not allow the magnetic beads to sit for more than 2 minutes before dispensing.
  - Before pipetting, shake or vortex the bottle to completely resuspend the Sequencing Magnetic Beads
1. Prepare a fresh Master Mix following Table1 for the number of samples to be processed, plus 10% more (e.g., if you have 10 samples, prepare Master Mix for 11). Add the following components to the reservoir.
  2. Add the following components to each well/or reagent reservoir based on table 1.
  3. Add 52  $\mu\text{L}$  premix the beads solution to each well of PCR plate or PCR tube (using 96-well PCR plates/tubes method) or each well of the microplate (using 96-Well Microplates).

**IMPORTANT!**

- Pipet up and down premix beads solution in a reagent reservoir until the solution is homogeneous before dispensing.
- Do not allow the magnetic beads to sit for more than 5 minutes before dispensing.

Table 1

| Component              | One well (10 $\mu\text{L}$ reaction volume) |
|------------------------|---|
| Sequencing Clean Beads | 7 $\mu\text{L}$                             |
| Ultrapure Water        | 45 $\mu\text{L}$                            |
| Total                  | 52 $\mu\text{L}$                            |

**B. Sample Processing**

1. After cycle sequencing is complete, remove the seal or cap and add 75  $\mu\text{L}$  **Ultrapure Water to each well/tube** and mix well to make a sequencing solution.
2. Aspirate 10  $\mu\text{L}$  sequencing solution to each well of plate or PCR tube in premix beads solution in step A3. (\*\* if necessary, centrifuge at 2500 rpm for 30 seconds to bring all contents to the bottom)
3. Vortex according to table 2 or table 3

Table 2

| 96-well PCR plates/PCR tubes |          |            |
|------------------------------|----------|------------|
| Vortexer                     | Speed    | Time       |
| Eppendorf MixMate            | 2000 rpm | 10 minutes |
| Mix-3000 Smart Mixer         | 2000 rpm | 5minutes   |

**IMPORTANT!**

The vortex step is critical to get the best results. We strongly recommend the use of the above vertex mixers and vortex conditions. Other compatible vortex mixers may be used. However, the vortex mixer must meet the following specificities: Orbit  $\geq 1.5\text{mm}$  and minimum speed  $\geq 2000$  rpm) and the vortex condition such as Speed and time has to be optimized.

Table 3

| 96-Well Microplates                                  |         |           |
|--|---------|-----------|
| Vortexer   | Speed   | Time      |
| Fisher Scientific™ Microplate Advanced Vortex Mixers | 800 rpm | 3 minutes |



|   |         |           |
|---|---------|-----------|
| OHAUS Microplate Vortex Mixers  | 800 rpm | 3 minutes |
| <p><b>IMPORTANT!</b><br/>The user can use other compatible vortex mixers. However, the mixing condition should be optimized, such as time and Speed, and the mixer should be Orbit <math>\geq 3</math> mm-4 mm, Speed <math>\geq 800</math> rpm</p> |         |           |

4. After vortexing, place the sample plate or PCR tube on the magnetic separation plate for 30 seconds or until the solution is clear.
  5. Centrifuge at 2500 rpm for 30 seconds to remove bubbles
  6. Place the sample plate or PCR tube on the magnetic separation plate for 30 seconds or until the solution is clear.
- C. Prepare the reaction plate for the capillary sequencing instruments. (**IMPORTANT!** Do not heat or use formamide with the sample.)

|                              |                    |  |
|------------------------------|--------------------|--|
|                              |                    |  |
| 96-well PCR plates/PCR tubes | Direct injection   | <ol style="list-style-type: none"> <li>1. Remove the plate from the magnetic rack and place a septum on the plate.</li> <li>2. Place a septum on the plate</li> <li>3. and load the reaction plate in the sequencer.</li> <li>4. Choose a BDX run module specified in the BigDye XTerminator™ Purification Kit User Guide (Pub. No. 4374408).</li> <li>5. Modify the run module by : <ul style="list-style-type: none"> <li>• adjusting injection to 25 seconds</li> <li>• adjusting injection voltage to 1200 v</li> </ul> </li> </ol> <p>The original Bigdye X-terminator Purification Kit run modules are available at <a href="http://www.thermofisher.com/sangerpatches">www.thermofisher.com/sangerpatches</a>.</p> <ol style="list-style-type: none"> <li>6. Start the electrophoresis run</li> </ol> |
|                              | Indirect injection | <ol style="list-style-type: none"> <li>1. Transfer 20-40 <math>\mu</math>L of the supernatant to a new plate, then place a septum on the plate. (** Make sure there is no bubble in the well. If necessary, centrifuge at 2500 rpm for 30 seconds to remove the bubble.)</li> <li>2. Place a septum on the plate</li> <li>3. load the reaction plate in the sequencer.</li> <li>4. Choose an appropriate run module</li> <li>5. Modify the run module by : <ul style="list-style-type: none"> <li>• adjusting injection to 25 seconds</li> <li>• adjusting injection voltage to 1200 v</li> </ul> </li> <li>6. Start the electrophoresis run</li> </ol>  |
| 96-Well Microplates          | Indirect injection | <ol style="list-style-type: none"> <li>1. Transfer 20-40 <math>\mu</math>L of the supernatant to a new plate, then place a septum on the plate. (** Make sure there is no bubble in the well. If necessary, centrifuge at 2500 rpm for 30 seconds to remove the bubble.)</li> <li>2. Place a septum on the plate</li> <li>3. load the reaction plate in the sequencer.</li> <li>4. Choose an appropriate run module</li> <li>5. Modify the run module by : <ul style="list-style-type: none"> <li>• adjusting injection to 25 seconds</li> <li>• adjusting injection voltage to 1200 v</li> </ul> </li> <li>6. Start the electrophoresis run</li> </ol>  |



**D. Troubleshooting**

| Problem                   | Probable cause  | Suggestion  |
|---------------------------|---|---|
| Dye Blobs (Dye artifacts) | Too much Dye  | The protocol is optimized for 2 µl or less of ABI BigDye Terminator v3.1. If possible, use 2 µl or less. or increase the amount of the magnetic beads.              |
|                           | Insufficient DNA template in the reaction                       | Increase DNA template concentration.  |
|                           | Incomplete removal of Dye                                       | <ul style="list-style-type: none"> <li>Optimize vortex time and Speed.</li> <li>Use the right vortex mixer.</li> <li>Use the right PCR tube or PCR plate</li> </ul> |
|                           | The magnetic beads are not properly suspended during dispensing | Thoroughly resuspend the magnetic beads before using them.  |
| Weak signal               | Improper reaction conditions                                    | Ensure a control sequencing reaction is performed during each thermocycling procedure and optimize reaction conditions if necessary.                                |
|                           | Injection time too short  | Increase injection time to 35-40 seconds.   |
|                           | Extension product concentration is too low.                     | Increase DNA template concentration.  |
|                           | Vortex time is too long, or vortex speed is too fast            | <ul style="list-style-type: none"> <li>Optimize vortex time and Speed.</li> <li>Use the right vortex mixer.</li> <li>Use the right PCR tube or PCR plate</li> </ul> |

**Related Products**

| <b>Products and Catalog Number</b>                           |  |
|--|--|
| <b>Genomic DNA and RNA Purification</b>                      |  |
| One-Step Mammalian Cell DNA Purification Kit, Cat. No. AA101 | One-Step Saliva Viral RNA-DNA Purification Kit, Cat. No. AR101 |
| Cell-Free DNA Purification Kit, Cat. No. AC101               | Bone-Teeth DNA Purification Kit, Cat. No. AB101                |
| One-Step FFPE & FNA DNA purification Kit, Cat. No. AJ-101    | Rootless Hair DNA Purification Kit, Cat. No. AD101             |
| One-Step Bacteria DNA Purification Kit, Cat. No. AE101       | One-Step Buccal Cell DNA Purification Kit, Cat. No. AG101      |
| One-Step Blood DNA Purification Kit, Cat. No. AF101          | One-Step Touch DNA Purification Kit, Cat. No. AS101            |
| One-Step Fungi & Yeast DNA Purification Kit, Cat. No. AL101  | Sexual Assault Casework DNA Purification Kit, Cat. No. AT101   |
| One-Step Insect DNA Purification Kit, Cat. No. AM101         | One-Step Fingerprint DNA Purification Kit, Cat. No. AZ101      |
| One-Step Mouse Tail DNA Purification Kit, Cat. No. AN101     | One-Step Dandruff DNA Purification Kit, Cat. No. AAA101        |
| One-Step Plant DNA Purification Kit, Cat. No. AQ101          | Quick mRNA Purification Kit, Cat. No. MMS101                   |
| <b>DNA &amp; RNA Sample Preparation</b>                      |  |
| One-Step NGS Cleanup Kit, Cat. No. AO101                     | One-Step DNA-RNA Removal Kit, Cat. No. CA103                   |
| One-Step RNA Removal Kit, Cat. No. AU101                     | One-Step DNA/RNA Cleanup Kit, Cat. No. AH101                   |
| One-Step PCR Cleanup Kit, Cat. No. AP101                     | One-Step Sequencing Cleanup Kit, Cat. No. AI101                |
| Quick Oligo-DNA Conjugation Kit, Cat. No. CA101              | One-Step Fluorescent Labeling Cleanup Kit, Cat. No. AK101      |
| One-Step DNA-RNA Removal Kit, Cat. No. AV101                 | One-Step Single-Stranded DNA Removal Kit, Cat. No. AW101       |
| One-Step PCR Inhibitor Removal Kit, Cat. No. AX101           | Pure Miniprep Plasmid DNA Purification Kit, Cat. No. AY101     |